Thermoanalytical and Starch Content Evaluation of Cassava Bagasse as Agro-Industrial Residue

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ABSTRACT

Starch nutritional fractions as well as thermal properties and other analysis are essential for food and industrial application. Cassava bagasse is an important agro-industrial residue and its starch content was evaluated using two alternative methods. Thermal characterization and microscopy analyses helped to understand how hydrolysis digests starchy fraction of cassava bagasse. The melting point of cassava starch occurred at 169.2°C. Regarding TG analyses, after moisture content, there were observed two main mass losses for all samples. Results suggest hydrolysis carried out using enzyme is less effective in order to convert total starch content in cassava bagasse. However, using sulfuric acid, fibers are affected by analyses conditions.

Key words: amylase, cassava, hydrolysis, thermal analysis

INTRODUCTION

Starch market comes growing in the last years, leading to the search of products with specific characteristics that support the requirements of the industry. The modified starch production is an alternative that is in continuous development to surpass one or more limitations of native starches and thus to increase the utility of this polymer for industrial applications (Lacerda et al., 2008; Leonel, Cereda and Roau, 1999; Rudnik et al., 2006; Wurzburg, 1986). Furthermore, there has been an increased exploitation of organic residues from various sectors of agriculture and industries over the past few decades. The use in biotechnological processes of crop residues such as bran, husk, bagasse, and fruit seeds regards as potential raw material in bioprocesses as they provide excellent substrates for the growth of microorganism supplying the essential nutrients to them (Pandey and Radhakrishnan, 1992; Pandey et al., 1994; Pandey and Soccol, 1998; Pandey et al., 2000). Their application in bioprocesses also offers advantages in bioremediation and biological detoxication of hazardous compounds (Ramachandran, 2000). Cassava (Manihot esculenta) ranks very high among crops that convert the greatest amount of solar energy and
CO$_2$ into carbohydrates per unit of area. Among
the starchy staples, cassava gives a carbohydrate
production which is about 40% higher than rice
and 25 % superior to corn, with the result that
cassava is the cheapest source of calories for both
human nutrition and animal feeding (Lacerda et
al., 2008). Normally, 69% of the starches derived
from the cassava are destined to nourishing use, in
meat products, pastas, desserts, breads, biscuits,
soups and candies. It has the remarkable capacity
to adapt to various agro-ecological conditions. It is
also considered as a low-risk crop (Pandey et al.,
2000). Cassava production is transformed into
flour and starch, generating US$ 600 million in
flour and 150 million in starch. During starch
extraction from cassava root, Leonel, Cereda and
Roau (1999), highlighted that the amount of solid
residues (mainly bagasses) generated is very great.
Currently, bagasse is being donated to farmers for
animal feeding, disposed in surrounding
environment of processing units or sold for US$ 5
per ton. The knowledge of starch content in
cassava bagasse is very important once this residue
can be used in further several industrial
applications being a high value-added material.
This study was performed to evaluate two usual
methods for starch content determination in
starchy material as well structural characterization
using thermal analysis.

MATERIALS AND METHODS

Samples
Cassava bagasse was received from a Starch
Industry (Comercial Agricola Anhumai) located in
the Northwest Region of Parana State-Brazil. The
material was dried and screened in order to obtain
particles < 1mm. α-amylase, from Bacillus
licheniformis (Termamyl 240L), was acquired
from Novozymes (Bagsvaerd, Denmark). Other
reagents (i.e. sulfuric acid) were of analytical
grade.

Hydrolysis
In order to solubilize starch content of cassava
bagasse, it was adopted chemical and enzymatic
hydrolyses as following description. For both
situations, the experiments were conducted three
times.

Chemical Hydrolysis
In this method, adapted from Woiciechovski
(2000), 2.0g of dry cassava bagasse (moisture
content of 10 %) were was added to 20.0 mL of
distilled water and 2.0 mL of sulfuric acid in an
erlenmeyer flask covered with aluminum foil
paper and rubber band. Hydrolysis reaction was
carried out in a water bath at approximately 98°C
during 20 min. After material cooling to room
temperature (about 25°C), the pH was adjusted to
7.0 using sodium hydroxide 0.5 mol/L. The total
volume was completed to 100mL with distilled
water in a calibrated volumetric flask (± 0.1 mL).
The aqueous medium was filtered in Whatman 42
filter paper and then an aliquot of 2.0 mL was
separated. Reducing sugar content of the filtered
solution, after a 1:10 (solution:water) dilution, was
estimated with a spectrophotometer SHIMADZU
UV-1601 at 540 nm following dinitrosalicilic
(DNS) method (Miller, 1959; Tastun, 1970). This
method involves spectrophotometric measurement
of reducing sugar liberated from a known soluble
starch medium by the action of amylase using
dinitrosalicylic acid reagent at pH 6.9 and room
temperature (Milanh et al., 2008).

Enzymatic hydrolysis
In order to hydrolyse studied matter using α-
amylase in excess, 1.0 g of cassava bagasse was
added to 25 mL of distilled water and 0.25 mL of
enzyme Termamyl 240L preparation in a 150.0
mL erlenmeyer flask. The pH was adjusted to 5.6
following enzyme data sheet instructions. After
covering erlenmeyer flask using aluminum foil
paper and rubber band, the sample was kept in a
water bath at 90-95°C during 25min. under
continuous stirring. Sample was removed from the
water bath and the pH of aqueous medium was
adjusted to 9.0 using sodium hydroxide 0.5 mol/L
in order to stop the enzyme action. The aqueous
medium was filtered in Whatman 42 filter paper
using a vacuum pump. Solid samples remaining on
paper filter was dried in an oven at 60°C during 2
days, and then kept in a desiccator over anhydrous
calcium chloride.

Thermal Analysis
Thermal analysis TG (Thermogravimetry), DTA
(Differential Thermal Analysis) and DSC
(Differential Scanning Calorimetry) curves were
recorded using a Shimadzu TG 60 and DSC 60, with synthetic air flowing at 100 mL/min, and a heating rate of 10°C/min. and with mass samples of about 6 mg. Alumina open sample holder and aluminum sealed crucibles were used for TG/DTA and DSC respectively. TG is used to measure the mass loss either as a function of time (isothermal) or dynamic temperature and controlled atmosphere (Mothé et al., 2006). Endothermic and exothermic changes in a DSC curve indicate events or reactions such as glass transition, gelatinization and melting, occurring during DSC analysis (Habitante et al., 2008).

Microscopy
Optical microscopy has been widely used by scientists and students as a useful tool to examine objects on a fine scale in order to get information relative to the morphology of the materials examined. After drying, samples of cassava bagasse untreated and treated by acid and enzyme were mounted on standard glass microscope slides. Microscopy analysis was carried out using an Olympus stereo microscope SZX9, with polarization filter and Cybernetic’s Cool Snap Pro Color camera. The photographs were identified and scaled using Image Pro Plus.

RESULTS AND DISCUSSION

Acid Hydrolysis
Acid hydrolysis has been used to modify starch for over 150 years (Duedahl-Olesen, Pedersen and Larsen, 2000) This process involves suspending starch in an aqueous solution of hydrochloric acid or sulfuric acid at certain temperatures. In the presence of a strong acid and heat, the glycosidic bonds between monosaccharides in a polysaccharide are cleaved. After hydrolysis using concentrated sulfuric acid as agent, from DNS method, it was obtained at 540 nm an optical density (OD) of 1.59 (± 0.13). Following calibration curve made for DNS reagent, glucose concentration of sample was estimated following as Eq.(1).

\[
RS (g/L) = (1.099OD+ 0.017) D
\]  

Where RS is related to reducing sugars concentration in g/L, OD is optical density at 540 nm, D is dilution of original hydrolysis solution and other values are related to calibration slope from standard curve. Thus, it was obtained a glucose concentration of 16.97 g/L (±0.66) in 100.0 mL completed with tap water. Considering initial moisture content in Table 1, actually it was converted 1.776g of cassava bagasse in dry basis, so each gram produced 0.95g/L of reducing sugars. Stoichiometrically, each 162g of starch, incorporates 18g of water during hydrolysis, producing 180g of glucose, thus there is a conversion factor equals to 1.11. In order to estimate starch content of studied sample it was used (Eq. (2)).

\[
SC_1 (\%) = \frac{(100) IGC}{1.11}
\]  

Where SC_1 is starch content, IGC is the initial glucose concentration and 1.11 is glucose-starch stoichiometric relation. Following the Eq. (2), cassava bagasse estimated starch content was 84.44% considering the studied sample. Acid hydrolysis is widely used as a method to determine starch (carbohydrate) content in a sample. However, degradation action is not specific to sugars and some fibrous matter is affected also. Based on reducing sugars, DNS method is not indicated when is needed to identify concentrations of specific carbohydrates with different molecular chain lengths (i.e. mono, di and trisaccharides).

Enzyme Hydrolysis
According to Duedahl-Olesen, Pedersen and Larsen (2000) and Swinkels (1985) α-amylases (Enzyme Classification 3.2.1.1) are endo-acting enzymes that randomly attack the internal α-D-(1.4) O-glycosidic linkages of starch except for those adjacent to the ends of the substrate and those in the vicinity of branch points. The end products are α-limit dextrins, which are branched saccharides not prone to further hydrolysis and malto-oligosaccharides of varying degrees of polymerization (DP), characteristic of the particular enzyme. Basically, starch content in this method was evaluated as difference among soluble fraction after hydrolysis and non soluble remaining material. Using enzyme bacterial α-amylase in excess, starch is converted into soluble fractions of maltose and other shorter chains of glucose units also known as dextrins. After drying the remaining material, final mass of 0.102g (±0.016) was observed. Considering moisture contents of untreated and hydrolyzed sample
showed in Table 1, starch content in dry basis was calculated by the (Eq.3).

$$SC_2 = (100) \times FM - 0.89$$  \hspace{1cm} (3)

Where $SC_2$ is estimated as starch content in sample, $FM$ is final mass or material mass remained on filter paper and 0.89 is related to moisture content. Cassava bagasse sample studied by enzyme hydrolysis has approximately 78.80% of starch. It is important to note that the use of enzymes can provide besides soluble fraction, the dietary fiber content. Actually, solid product from hydrolysis is mainly fiber and other compounds such as minerals and eventually lipids and others. Faithfull (1990), studying the determination of starch content in potatoes ($Solanum tuberosum$) by enzyme hydrolysis observed several disadvantages comparing to the acid hydrolysis procedure. The extraction and hydrolysis process is slower, especially using amyloglucosidase, enzyme activity may vary, reagents are more expensive, and complete hydrolysis is more difficult. Its main advantage is that it is specific, not affecting the cellulosic polysaccharides in the cell walls. It is important to mention that both methods studied are very quick, useful and low cost procedures. However they have experimental limitations.

**Thermal analysis**

Conventional plastics have large impact in increasing the environment’s pollution. Thus, the interest has turned towards novel partially and completely biodegradable polymers (Schlemmer, de Oliveira and Salles, 2007). Structural resistance of starches is a very important factor on biodegradable plastics studies and the knowledge regards on blending of starch and polymers. DSC endothermic profile curve, showed in Fig 1, is related to melting event of cassava starch presented in bagasse. The enthalpy required for the event was estimated as 129.16 J/g and melting point occurred at 169.36°C, confirming thermal event result observed previously by Siroth et al., (2000).

Initial mass losses temperatures are presented in Table 1 and all analyzed samples showed three main mass losses events. Raw cassava bagasse presented was the first one with 11.2%, the second 65.3% and the third, 19.8% of mass loss. At the end of the process remains about 1.3% as a residue.

**Figure 1 - DSC curve of dry cassava bagasse.**
Table 1 - Thermal degradation of cassava bagasse in untreated state and after hydrolysis process.

<table>
<thead>
<tr>
<th>Cassava bagasse</th>
<th>Untreated (a)</th>
<th>Submitted to acid treatment (b)</th>
<th>Submitted to enzyme treatment (c)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Moisture (%)</td>
<td>11.21</td>
<td>5.61</td>
<td>9.94</td>
</tr>
<tr>
<td>Total mass loss (%)</td>
<td>98.27</td>
<td>96.34</td>
<td>96.02</td>
</tr>
<tr>
<td>DTA peak 1 (ºC)</td>
<td>335.59</td>
<td>317.21</td>
<td>329.35</td>
</tr>
<tr>
<td>DTA peak 2 (ºC)</td>
<td>442.73</td>
<td>448.37</td>
<td>465.07</td>
</tr>
<tr>
<td>On-set 1 (ºC)</td>
<td>273.56</td>
<td>257.72</td>
<td>243.96</td>
</tr>
<tr>
<td>On-set 2 (ºC)</td>
<td>445.87</td>
<td>431.49</td>
<td>465.31</td>
</tr>
</tbody>
</table>

Cassava bagasse TG curve after acid treatment showed in Fig 2, presents the same three mass losses. The first 5.6%, second 65.7% and the third 20.9%. Finally, after α-amylase hydrolysis, cassava bagasse presented first loss of 9.94%, the second of 69.4% and the third of 13.8% of the total mass. The temperatures were 39.35 to 150, 246.30 and 458.07°C.

As described by LeVan (1998) in all TG curves observations, from 30°C to 150ºC, mass loss regards to the elimination of moisture content; the second loss occurs between 200 and 400°C represents the decomposition of hemicellulose and cellulose the third loss stage above 400ºC probably concerns to the decomposition of lignin and ash formation. The hemicellulose is less stable than the cellulose because its side chains and degrades before cellulose. The lignin degrades throughout hemicellulose and cellulose degradation processes (Shafizadeh and DeGroot, 1976). Comparing the DTA curves, the peak which represents lignin degradation above 400°C is shorter and less acute in the acid treated cassava bagasse than in raw or enzyme hydrolyzed. This variation occurred probably because acid treatment is able to degrade fiber which was not attacked in hydrolysis using enzyme.

![Figure 2 - TG curves of (a) untreated cassava bagasse, (b) acid treated cassava bagasse and (c) cassava bagasse after enzyme hydrolysis.](image)

The TG curve of raw cassava bagasse shows three stages of mass loss. The first stage between 30°C and 150°C is due to the loss of moisture content which represents 11.19% of the total mass. The next stage begins at 255.9°C and represents the decomposition of hemicellulose and cellulose bonds and, in addition, for untreated samples complete breakdown of starch (Aggarwal and Dollimore, 1998; Rudnik et al., 2006). Since next degradation (close to 340°C) does not occur in inert atmosphere, it was supposed that the oxidation of the partially decomposed matter starts TG curves of the acid treated cassava bagasse, shows that the amount of lignin decreased due to the reducing of the degradation peak’s. This variation can be explained by the strong and
randomic acid reaction, breaking more compounds than the enzymatic treatment, which cleaves only starchy material as observed in microscopy. DTA curve of raw cassava bagasse in Fig 3 shows the exothermic peak at 343.13°C which is associated to oxidation of cellulose into levoglucosan, water, carbon monoxide and carbon dioxide (Aggarwal and Dollimore, 1998).

![Figure 3 - DTA curves of (a) untreated cassava bagasse, (b) acid treated cassava bagasse and (c) cassava bagasse after enzyme hydrolysis.](image)

Compounds are generated from both cellulose and hemicellulose (hemicellulose is less stable). DTA peak at 446.53°C shows the lignin degradation, more specific by breaking (cleaving) carbon-carbon bonds. At the end of the process, 1.73% of residue still remains being related to ashes.

**Microscopy**

After microscopic analysis, it can be observed in Fig 4(a) high amount of free cassava starch with its characteristic rounded shape. Besides the presence of a lot of free granules, the untreated material also has fibers with high amount of starch bonded. As previously studied by Leonel, Cereda and Roau (1999) through microscopic observations it is possible to confirm the complete hydrolysis of free starch granules using amylolytic enzyme as showed in Fig 4(b) and the same result was observed using acid treatment. However, cell walls still remains using enzyme and after treatment, some gelatinized starchy material can be present after hydrolysis. Acid treatment can even destroy cell walls as showed in figure 4(c) thus, suggesting that there is not any starch remained after hydrolysis.

![Figure 4 - Photomicrograph of (a) untreated cassava bagasse, (b) acid treated cassava bagasse and (c) cassava bagasse after enzyme hydrolysis.](image)
CONCLUSIONS

Through microscopy analysis, it was observed that cassava bagasse hydrolyzed with enzyme α-amylase presented cell wall without severe structural changes; however, there still starchy matter in the cells alls even after hydrolysis. Chemical hydrolysis using sulfuric acid leaded to a material critical degradation of both starchy and fibrous matter and due to this action, probably there was not any starch remained. Thermal analysis helped to understand the characterization of all samples of cassava bagasse studied regarding mass losses behaviors as function of temperature increasing. Despite the fact of different starch contents among studied methods, there is a high amount of starch in samples. Thus, the disposal of cassava bagasse concerns on environmental problem and several potential losses of industrial and social opportunities by the use of this residue.

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